

A Short Review on Phytoconstituents from the Genera *Albizzia* and *Erythrina*

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Abstract

The genus *Albizzia* and *Erythrina* are the leading sources of phytoconstituents. The aim of this review is to sollicitude of the phytoconstituents from some medicinal plants. A total nine medicinal plants were studied and 121 chemical constituents along with structures have been reported here. *Erythrina burtii* consists of highest number of constituents.

Key words: Medicinal plant, Phytoconstituents, Flavonoids, Isoflavanones, Saponins, Sesquiterpenes, Flavan, Steroid, and Triterpenoids.

Introduction

Nature is a great source of medicinal plants and these plants are used as a traditional medicine for many years (Hussain *et al.*, 2010). One hundred and ten species are present as trees and shrubs in the genus *Erythrina* (Hussain *et al.*, 2016a, 2011). Among them, two species have been reviewed comprehensively in this review. *E. burtii* is a flowering and flat-topped tree (height: 3.5-18 m) growing in Ethiopia, Kenya, and Tanzania. *E. droogmansiana* is a single straight stem, soft wood and rounded crown tree (height: 20 m) extensively grown in Congo, Cameroon and Gabon, and used in the treatment of fever, hemorrhoids, and wound infection in locally. The genus *Albizzia* consists of 150 species extensively distributed in Africa, Asia, and South America. *Albizzia* species were used as traditional medicine in the treatment of anthelmintic, cough, diarrhea, insomnia, irritability, injuries, poor memories, rheumatism, scabies, stomach trouble, and wounds in Africa and China (Hussain *et al.*, 2016b). *A. anthelmintica* is a medium canopied tree (height: 8 m) with soft bark and unwrap spine. *A. lebeck* (Leguminosae) is an exposed deciduous tree (height:

12-21 m) that grows in over Bangladesh (Hussain *et al.*, 2008). *A. inundata* is a perennial tree and found in Argentina. *A. glaberrima* is a big tree having few flattened crown and used as a folk medicine in the treatment of anemia, blenorhagia, bilharzias, epilepsy, and liver complications in Cameroon and Nigeria. *A. coriaria* is a medicinal plant found in Uganda and used in the treatment eye diseases, jaundice, skin disease, sore throats, and syphilis. *C. zeyheri* (Family: Combretaceae) is a Tanzanian medicinal plant and applied for the management of different health consequences such as cancer, cough diarrhea, hypertension, and snakebite.

Reported phytoconstituents

A total nine medicinal plants have been studied and one hundred and twenty one (1-121) compounds were reported in this review as phytoconstituents. The studied medicinal plants are *Erythrina burtii*, *E. droogmansiana*, *Albizzia submidiata*, *A. anthelmintica*, *A. inundata*, *Spergularia marginata*, *Manikara rufula*, *A. lebeck*, *Ainsliaea yunnanensis*, *A. glaberrima*, *Combretum zeyheri*, *A. boromoensis*, *A. grandibracteata*, and *A. coriaria*.

Flavonoids

Erythrina genus is a pioneer source of flavonoids. There are twenty four phytoconstituents have been reported as flavonoids and pilocarpin from *Erythrina burtii* such as Burttinol A (1), Burttinol-A diacetate (2), Burttinol B (3), Burttinol-B acetate (4), Burttinol C (5), Burttinol-C diacetate (6), Eryvarin H (7), Eryvarin-H diacetate (8), Burttinol D (9),

Burttinol-D diacetate (10), 4-O-methylsigmoldin B (11), Abyssinone V (12), Abyssinone V methyl ether (13), Calopocarpin (14), Burttinne (15), Neurautenol (16), Bidwillon (17), Isobavachalcone (18), Erythrasinate (19), 7-O-methyluteone (20), Burttinonedehydrate (21), 8-Prenylluteone (22), 3-O-methylcalopocarpin (23), and genistein (24) (Figure 1) (Yenesew *et al.*, 2012, 1998, 2003).

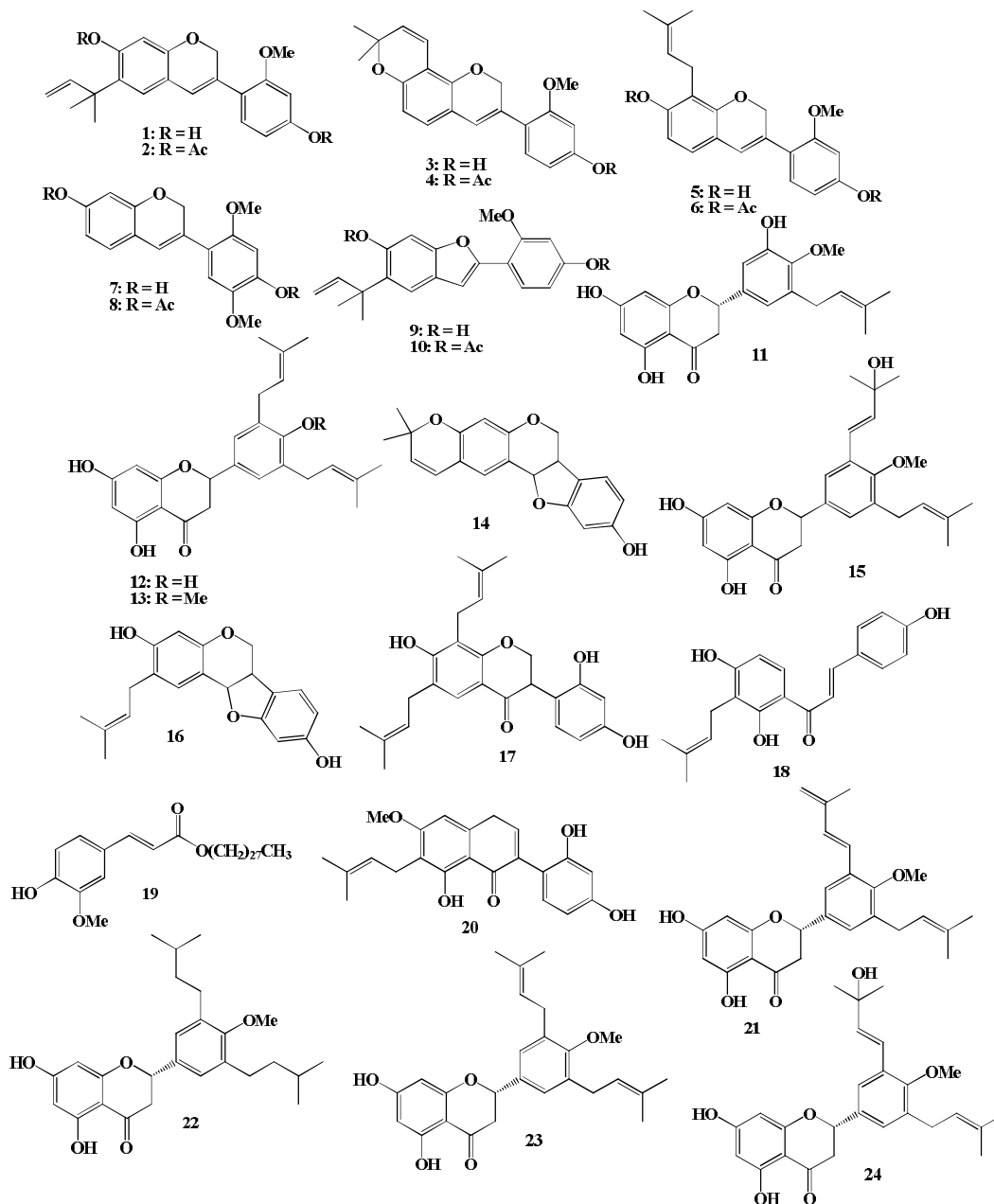


Figure 1. Flavonoids from *Erythrina burtii*.

Isoflavanones

The genus *Erythrina* (Family: Leguminosae) is a renowned source of isoflavanones and alkaloids. Ten isoflavanones were reported from the root bark of *Erythrina droogmansiana* for example 7,4'-Dihydroxy-2'-methoxy-3'-(3-methylbut-2-enyl)-isoflavanone (25), Sophoraisoflavanone A (26),

Erypoeigin D (27), Trihydroxy-8-(3-methylbut-2-enyl)-[6'',6''-dimethyl-pyrano(2'',3'', 4',5')]isoflavone (28), Isolupalbigenin (29), 5,7,2',4'-Tetrahydroxy-8,5'-di-(3-methylbut-2-enyl)-isoflavone (30), Erypstyrene (31), Phaseollidin (32), Cristacarpin (33), and Erystagallin A (34) (Figure 2) (Bedane *et al.*, 2017).

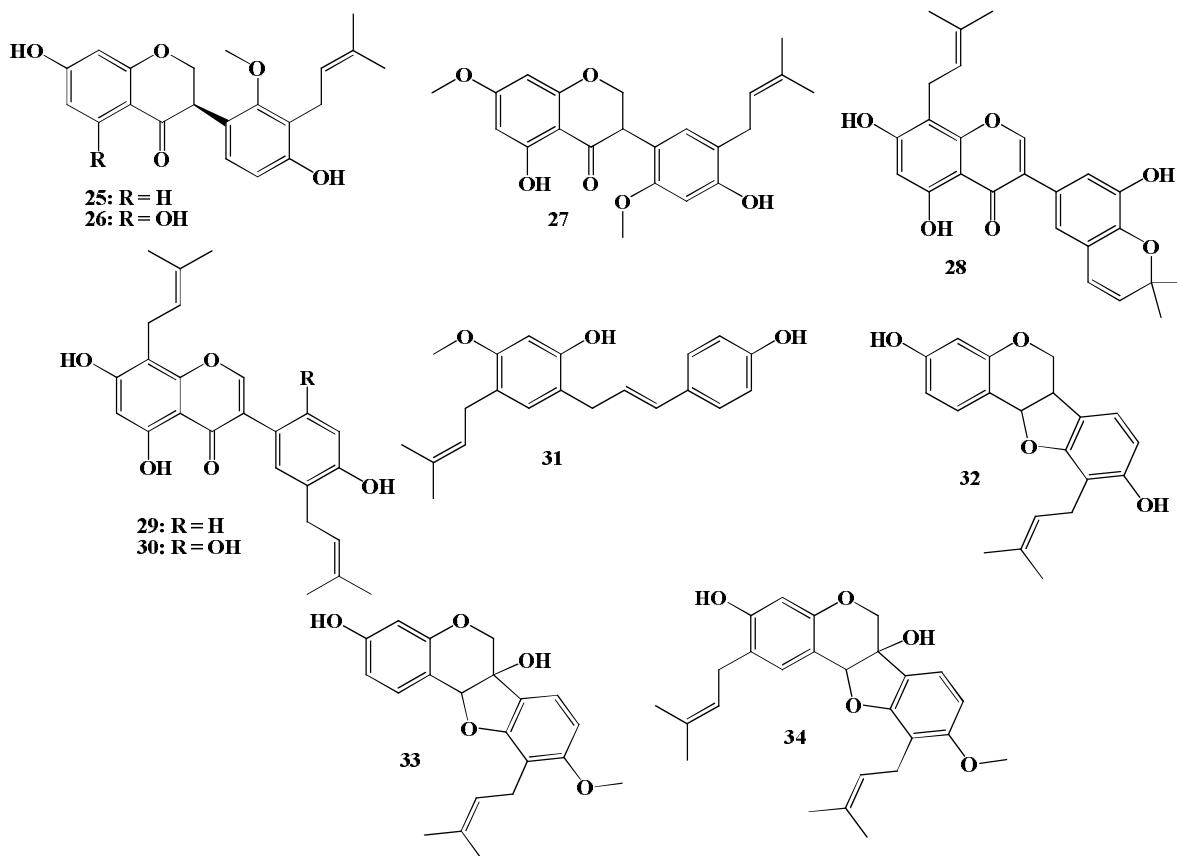


Figure 2. Isoflavanones from *Erythrina droogmansiana*.

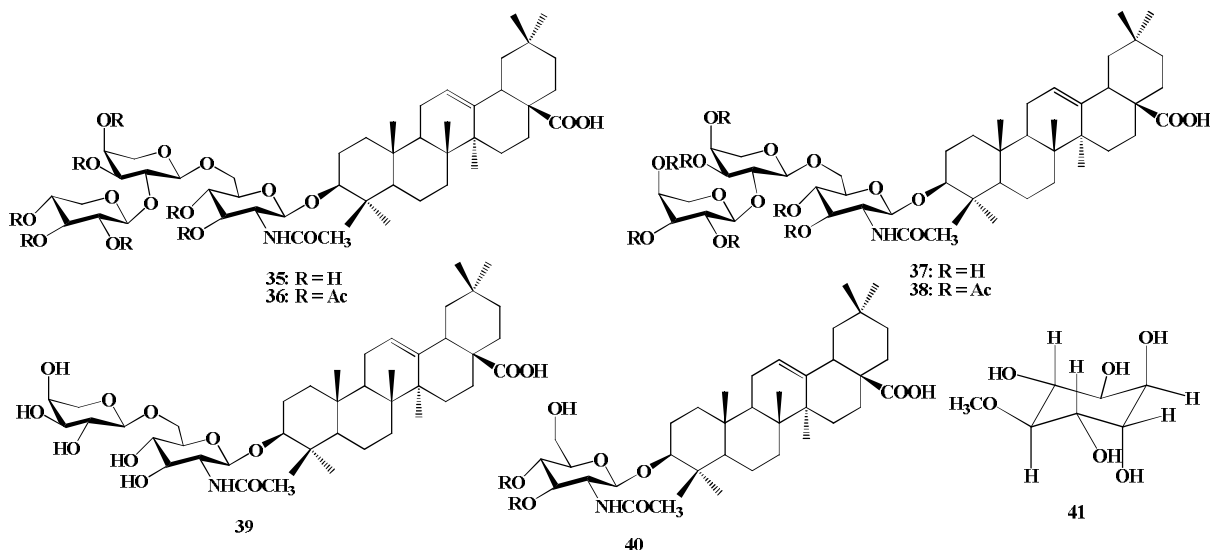
Saponins

Albizia subdimidiata, *A. anthelmintica*, *A. inundata*, *Spergularia marginata*, and *Manilkara rufula* are the key source of saponins. The reported saponins from these plants are 3-O-D-Xylopyranosyl-(1→2)-L-arabinopyranosyl-(1→6)-2-acetamido-2-deoxy-D-glucopyranosyl-oleanolic acid (35), 3-O-D-Xylopyranosyl-(1→2)-L-arabinopyranosyl-(1→6)-2-acetamido-2-deoxy-D-glucopyranosyl-oleanolic acid peracetate (36), 3-O-L-

Arabinopyranosyl-(1→2)-L-arabinopyranosyl-(1→6)-2-acetamido-2-deoxy-D-glucopyranosyl-oleanolic acid (37), 3-O-L-Arabinopyranosyl-(1→2)-L-arabinopyranosyl-(1→6)-2-acetamido-2-deoxy-D-glucopyranosyl-oleanolic acid peracetate (38), 3-O-L-Arabinopyranosyl-(1→6)-2-acetamido-2-deoxy-D-glucopyranosyl-oleanolic acid (39), 3-O-2-Acetamido-2-deoxy-D-glucopyranosyl-oleanolic acid (40), O-Methyl-cyclitol (41), 3-O-[α-L-Arabinopyranosyl-(1→6)]-2-acetamido-2-deoxy-β-

D-glucopyra-nosyl-oleanolic acid (**42**), 3-O-[α -L-Arabinopyra-nosyl-(1 \rightarrow 6)]-2-acetamido-2-deoxy- β -D-glucopyra-nosyl-echinocystic acid (**43**), 3-O-[α -L-Arabinopyra-nosyl-(1 \rightarrow 2)- α -L-arabinopyra-nosyl-(1 \rightarrow 6)]-2-acetamido-2-deoxy- β -D-glucopyra-nosyl-acacic acid lactone (**44**), 3-O-[β -D-xylopyranosyl-(1 \rightarrow 2)- α -L-arabinopyranosyl-(1 \rightarrow 6)]-2-acetamido-2-deoxy- β -D-glucopyranosyl-acacic acid lactone (**45**), 3-O-[α -L-arabinopyranosyl-(1 \rightarrow 2)- α -L-arabinopyranosyl-(1 \rightarrow 6)]- β -D-glucopyra-nosyl-oleanolic acid (**46**), 3-O-[β -D-xylopyra-nosyl-(1 \rightarrow 2)- α -L-arabinopyra-nosyl-(1 \rightarrow 6)]- β -D-glucopyra-nosyl-oleanolic acid (**47**), 3-O-[β -D-glucopyra-nosyl-(1 \rightarrow 2)]- β -D-glucopyra-nosyl-oleanolic acid (**48**), 3-O-[α -L-arabinopyra-nosyl-(1 \rightarrow 2)- α -L-arabinopyra-nosyl-(1 \rightarrow 6)]- β -D-glucopyra-nosyl-(1 \rightarrow 2)]- β -D-glucopyra-noside-echinocystic acid (**49**), 3-O-[β -D-xylopyra-nosyl-(1 \rightarrow 2)- α -L-arabinopyra-nosyl-(1 \rightarrow 6)]- β -D-glucopyra-nosyl-(1 \rightarrow 2)]- β -D-glucopyra-noside-echinocystic acid (**50**), 3-O-[β -D-glucopyra-nosyl-(1 \rightarrow 3)]-[α -L-arabinopyra-nosyl-(1 \rightarrow 2)]-[α -L-arabinopyra-nosyl-(1 \rightarrow 6)]-2-acetamido-2-deoxy- β -D-glucopyra-nosyl-echinocystic acid (**51**) 3-O-[α -L-arabinopyra-nosyl-(1 \rightarrow 2)]-[α -L-arabinopyranosyl-

(1 \rightarrow 6)]-2-acetamido-2-deoxy- β -D-glucopyra-nosyl-echinocystic acid (**52**), 3-O-[α -L-Arabinopyranosyl-(1 \rightarrow 6)]-2-acetamido-2-deoxy- β -D-glucopyra-nosyl-echinocystic acid (**53**), 3-O- β -D-glucuronopyra-nosyl-echinocystic acid 28-O- α -L-arabinopyra-nosyl-(1 \rightarrow 2)- α -L-rhamnopyra-nosyl-(1 \rightarrow 3)- β -D-xylopyra-nosyl-(1 \rightarrow 4)- α -L-rhamnopyranosyl-(1 \rightarrow 2)- α -L-arabinopyra-nosyl ester (**54**), 3-O- β -D-glucopyra-nosyl-(1 \rightarrow 3)- β -D-glucuronopyra-nosyl echinocystic acid 28-O- α -L-arabinopyra-nosyl-(1 \rightarrow 2)- α -L-rhamnopyra-nosyl-(1 \rightarrow 3)- β -D-xylopyra-nosyl-(1 \rightarrow 4)- α -L-rhamnopyra-nosyl-(1 \rightarrow 2)- α -L-arabinopyra-nosyl ester (**55**), 3-O- β -D-glucopyra-nosyl-(1 \rightarrow 4)-3-O-sulfate- β -D-glucuronopyra-nosyl echinocystic acid 28-O- α -L-arabinopyra-nosyl-(1 \rightarrow 2)- α -L-rhamnopyra-nosyl-(1 \rightarrow 3)- β -D-xylopyra-nosyl-(1 \rightarrow 4)- α -L-rhamnopyra-nosyl-(1 \rightarrow 2)- α -L-arabinopyra-nosyl ester (**56**), 3-O- β -D-glucopyra-nosyl-(1 \rightarrow 4)- β -D-glucuronopyra-nosyl-21-O-acetyl acacic acid (**57**), and Mi-saponin C (**58**) (Figure 3) (Kader *et al.*, 2001; Runyoro *et al.*, 2015; Zhang *et al.*, 2011; Carpani *et al.*, 1989; Pertuit *et al.*, 2017; Vieira *et al.*, 2017).



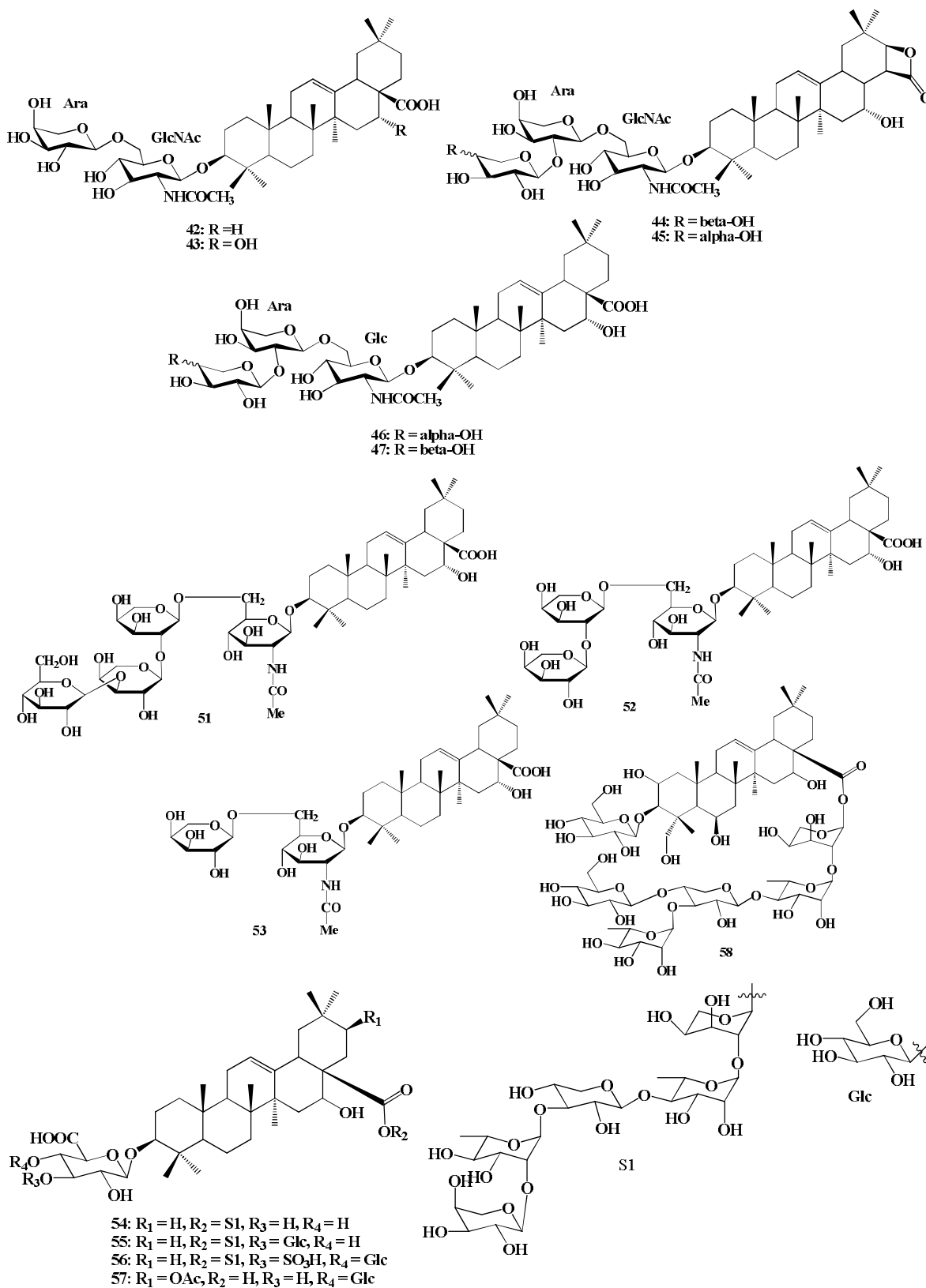


Figure 3. Reported saponins from medicinal plants.

Sesquiterpenes

Aromatic medicinal plant *Albizzia lebeck* and *Ainsliaea yunnanensis* are the principle source of sesquiterpenes. The elucidated sesquiterpenes from these plant are Benzyl-1-O- β -D-glucopyranosidem (59), Benzyl-6-O- α -L-arabinopyranosyl- β -D-glucopyranoside (60), Linalyl- β -D-glucopyranoside (61), Linalyl-6-O- α -L-arabinopyranosyl- β -D-glucopyranoside (62), (2E)-3,7-Dimethylocta-2,6-dienoate-6-O- α -L-arabinopyranosyl- β -D-glucopyranoside (63), Glycoside 1-O-[6-O- α -L-arabinopyranosyl- β -D-glucopyranoside]-(2E, 6E)-farnesol (64), *n*-Hexyl- α -L-arabinopyranosyl-(1 \rightarrow 6)- β -D-glucopyranoside (65), *n*-Octyl- α -L-arabinopyranosyl-(1 \rightarrow 6)- β -D-glucopyranoside (66), 2,3-Dihydroxy-2,3-dihydrosqualene (67), Ethyl fructofuranoside (68), Yunnanolides A (69), Yunnanolides B (70), Yunnanolides C (71), Yunnanolides D (72),

Yunnanolides E (73), Yunnanolides F (74), Yunnanolide G (75), Yunnanolides H (76), Yunnanolides I (77), Pertyolide C (78), Diaspanolide A (79), Diaspanolide B (80), 1 α -Hydroxy-3-O-isovalerate zaluzanin C (81), Tetrahydrodehydrozaluzanin C (82), Dihydrozaluzanin C (83), Zaluzanin C (84), Isoamberboin (85), 11b,13-Dihydro-3-epizaluznin C (86), and 4b,15,11b,13-Tetrahydrozaluzanin C (87) (Figure 4 and 5) (Massarani *et al.*, 2016; Fang *et al.* 2017).

Flavan and Steroids

A bunch of flavan and steroids were isolated from the *Albizzia glaberrima* such as (+)-(2R,3S,4R)-3',4',7-trihydroxy-4-methoxy-2,3-trans-flavan-3,4-trans-diol (88), (+)-Mollisacacidin (89), (+)-Fustin (90), Butin (91), Chondrillasterol (92), and Chondrillasterone (93) (Figure 6) (Fotso *et al.*, 2017).

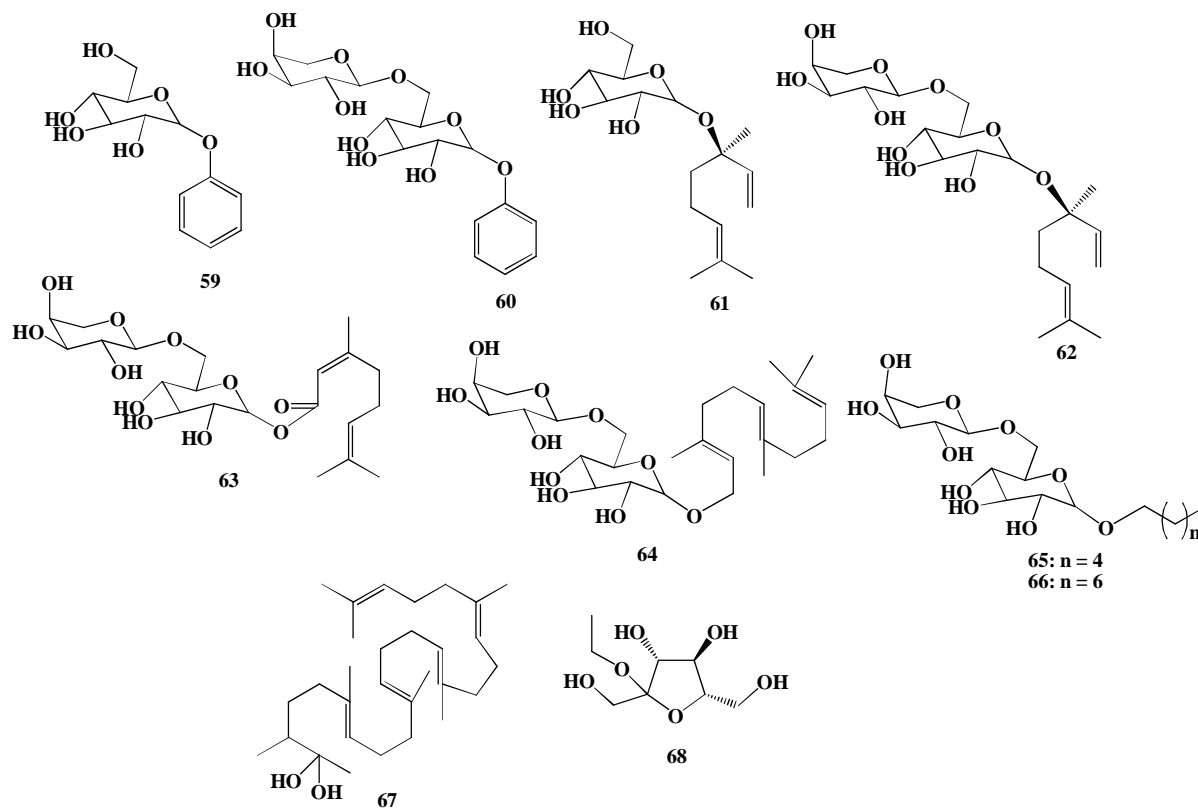


Figure 4. Sesquiterpenes from *A. lebeck*.

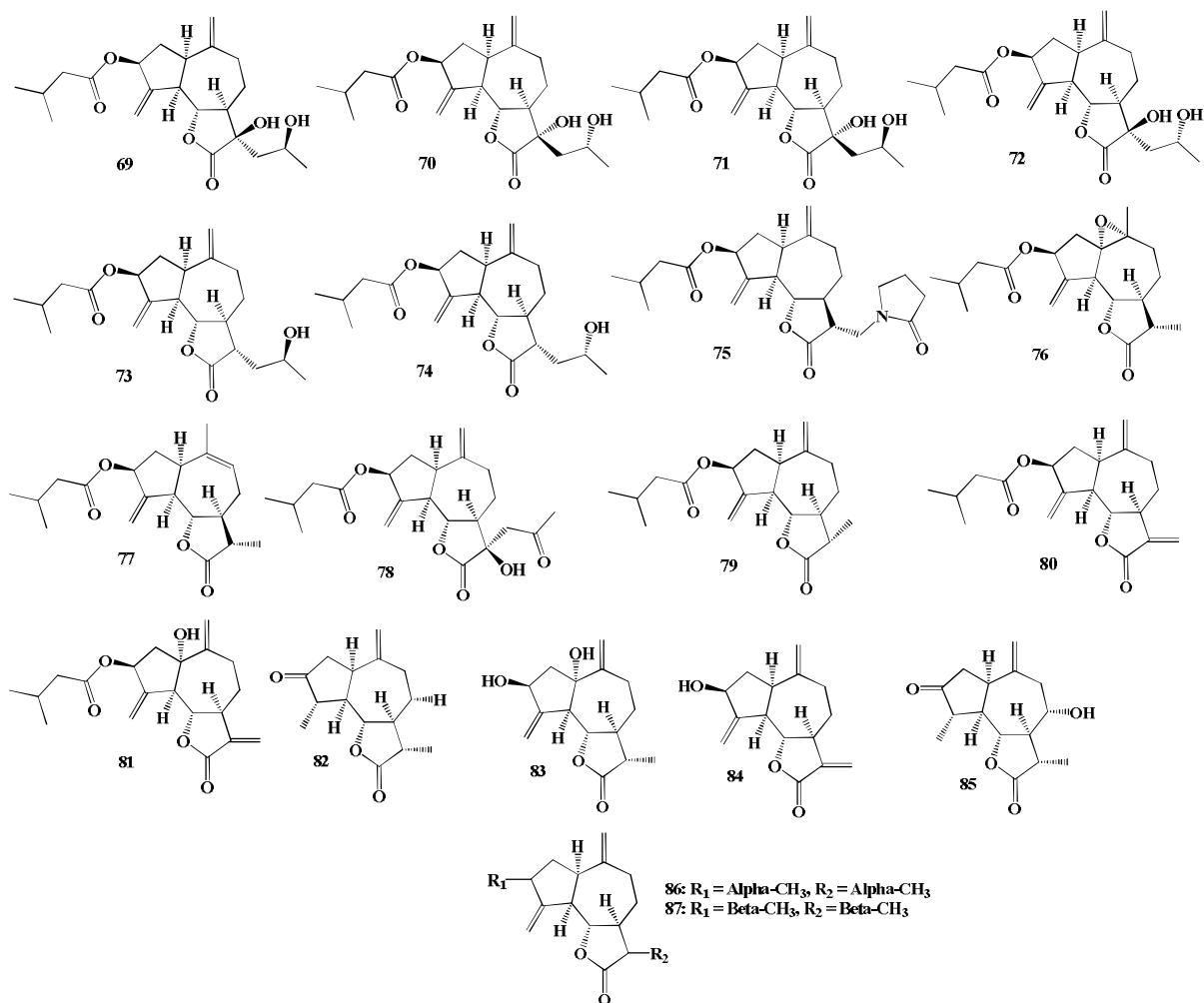
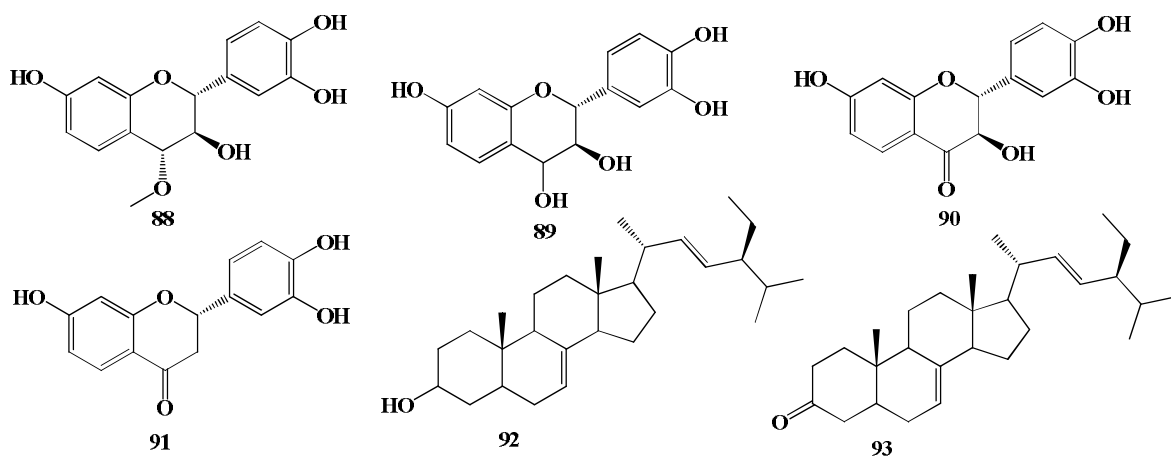


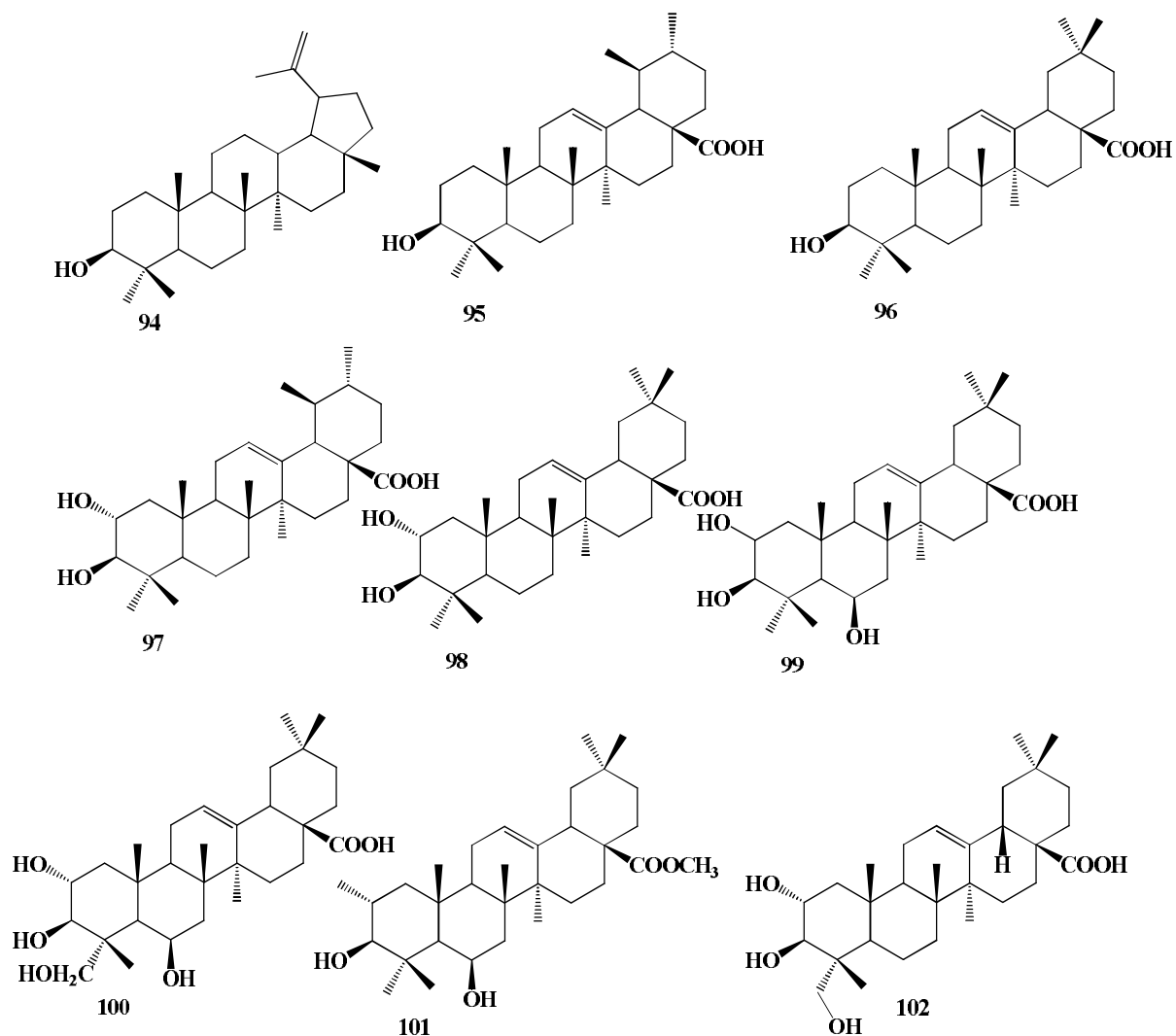
Figure 5. Isolated sesquiterpenes

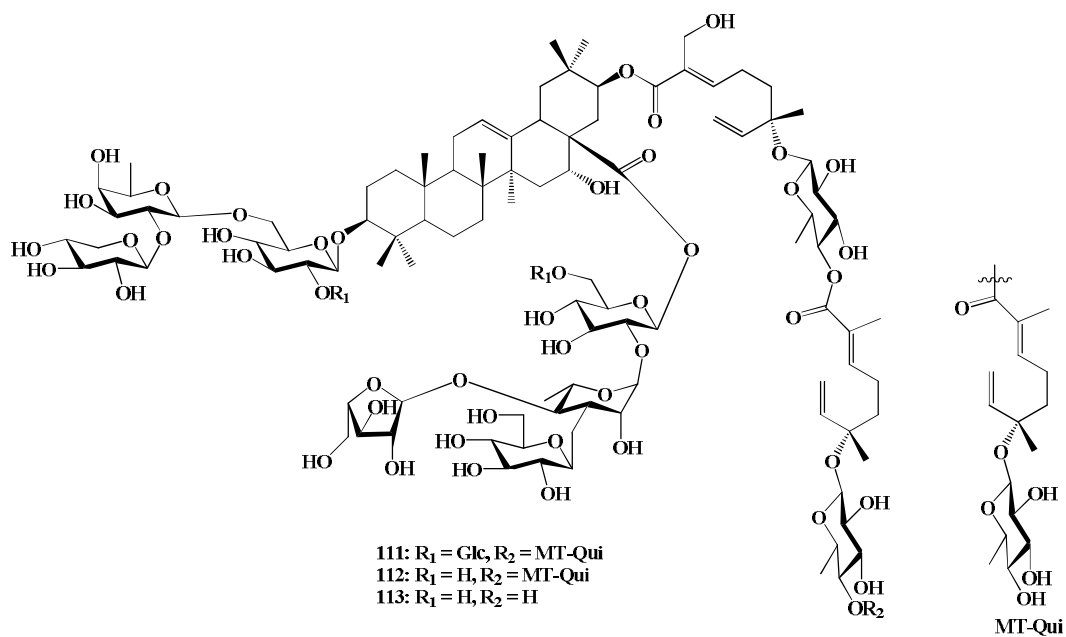
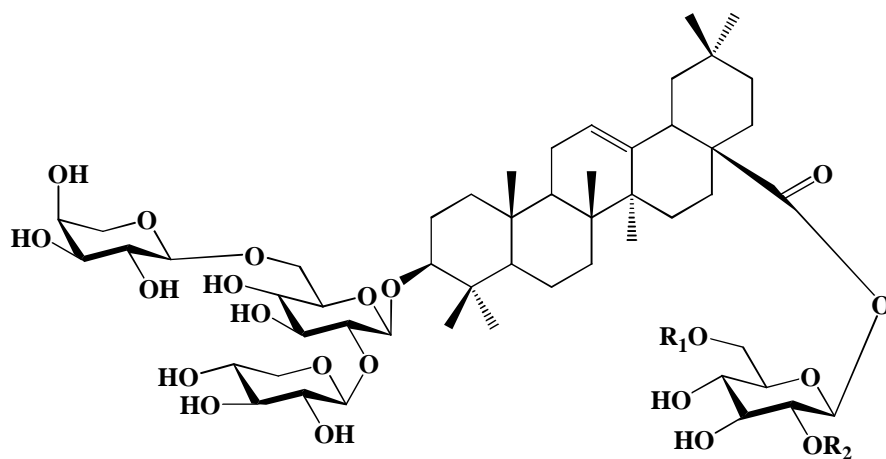
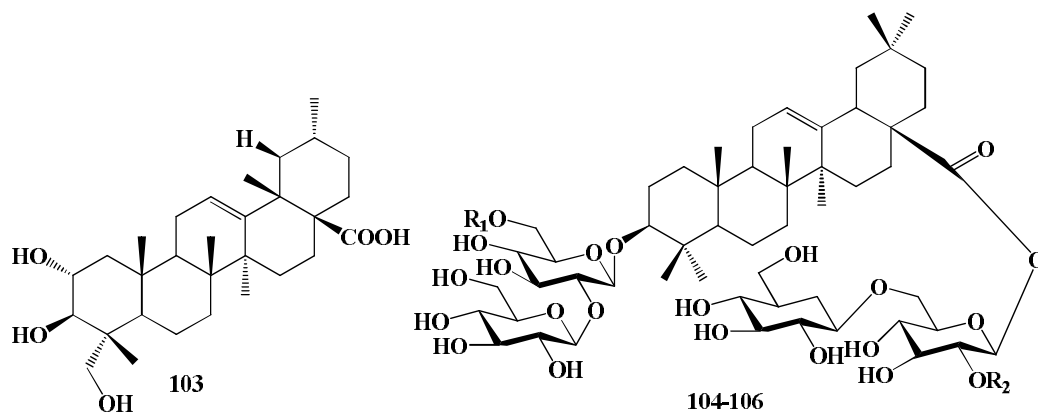
Figure 6. Flavan and steroids from *Albizzia glaberrima*.

Triterpenoids

A lots of triterpenoids have been derived from *Combretum zeyheri*, *A. glaberrima*, *A. boromoensis*, and *A. grandibracteata* for example Lupeol (94), Ursolic acid (95), Oleanolic acid (96), Maslinic acid (97), 2 α ,3 β -Dihydroxy-urs-12-en-28-oic acid (98), 6 β -Hydroxymaslinic acid (99), Terminolic acid (100), Methylsumaresinolate (101), Arjunolic acid (102), Asiatic acid (103), Glaberrimoside A (104),

Glaberrimoside B (105), Glaberrimoside C (106), Boromoenoside A (107), Boromoenoside B (108), Boromoenoside C (109), Boromoenoside D (110), Gummiferaosides D (111), Gummiferaosides E (112), Julibroside J₈ (113), Grandibracteoside A (114), Grandibracteoside B (115), and Grandibracteoside C (116) (Figure 7) (Runyoro *et al.*, 2013; Note *et al.*, 2016, 2015; Simo *et al.*, 2017; Krief *et al.*, 2005).





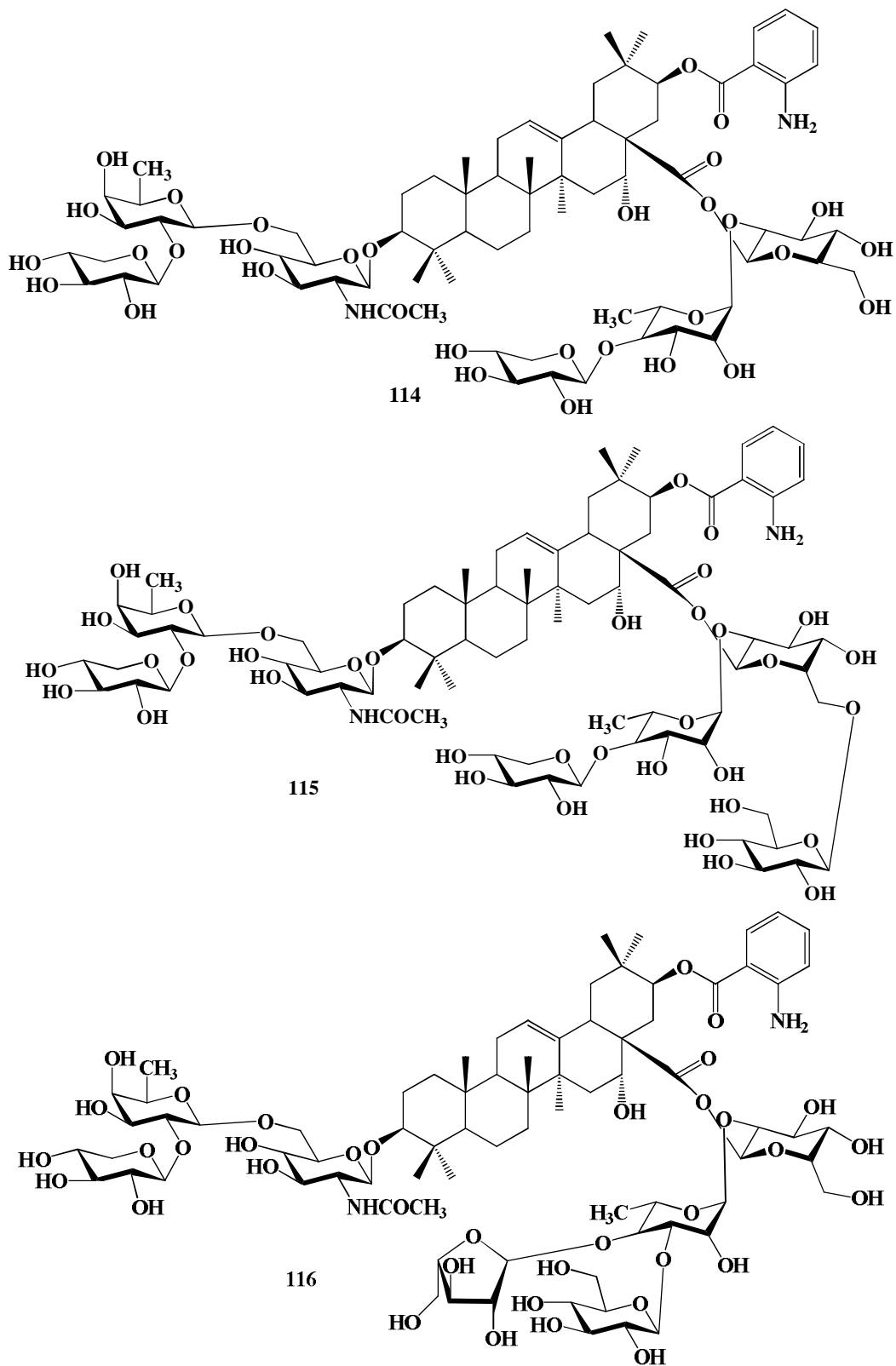


Figure 7. Reported triterpenoids.

Miscellaneous

A total six molecules for examples Lupeol (94), Lupenone (117), Betulinic acid (118), Acacic acid lactone (119), (+)-Catechin (120), and Benzyl alcohol (121) were isolated with chemical structures

from *Albizzia coriaria* (Figure 8) (Byamukama *et al.*, 2015).

Biological properties

The reported phytoconstituents showed lots of biological properties that are given in table 1.

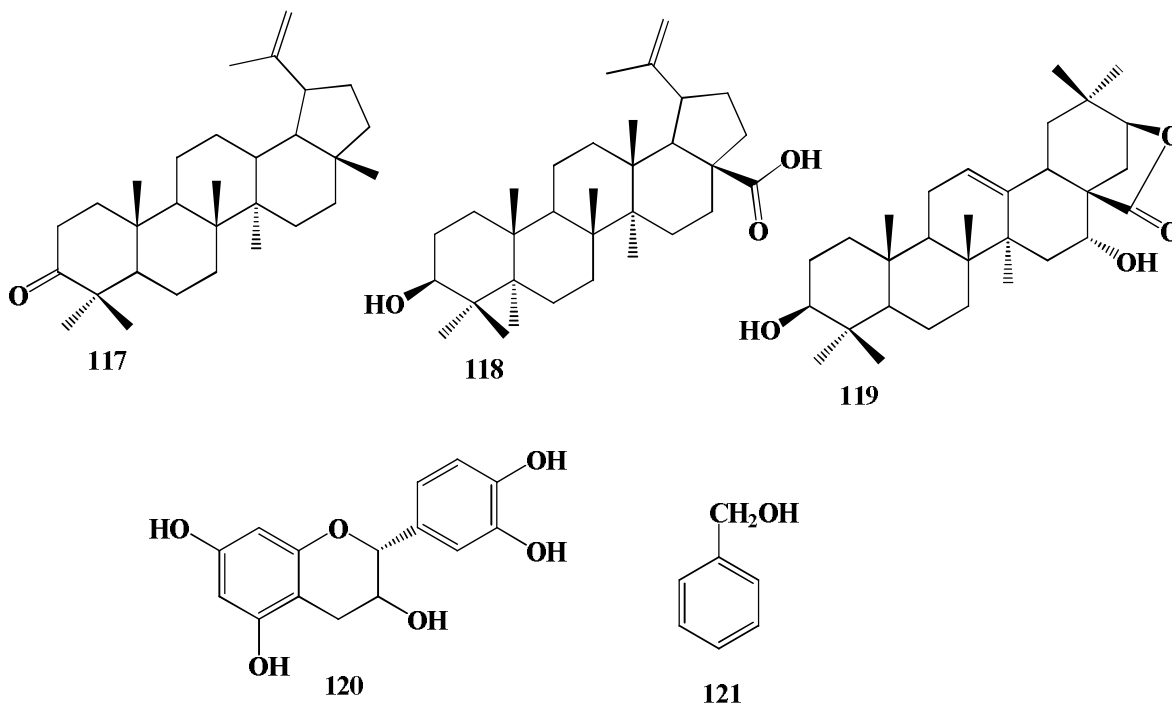


Figure 8. Miscellaneous compounds from *Albizzia coriaria*.

Table 1. Biological properties of the reported phytoconstituents.

Molecules	Biological properties	Ref.
1-24	Antiplasmodial and DPPH free radical scavenging	Yenesew <i>et al.</i> , 2012, 1998, 2003
25-34	DPPH free radical scavenging	Bedane <i>et al.</i> , 2017
34-57	Cytotoxic	Kader <i>et al.</i> , 2001; Runyoro <i>et al.</i> , 2015; Zhang <i>et al.</i> , 2011; Carpani <i>et al.</i> , 1989; Pertuit <i>et al.</i> , 2017
58	Anti-trichomonal	Vieira <i>et al.</i> , 2017
69-87	Inhibitory effect against nitric oxide	Fang <i>et al.</i> 2017
88-93	Cytotoxic	Fotso <i>et al.</i> , 2017
94-106	Anticandida and cytotoxic	Runyoro <i>et al.</i> , 2013; Note <i>et al.</i> , 2016
107-110	Inhibitory effect	Note <i>et al.</i> , 2015
111-113	Pro-apoptotic activity (Cytotoxic)	Simo <i>et al.</i> , 2017
114-116	Inhibitory activity	Krief <i>et al.</i> , 2005
117-121	Antimicrobial	Byamukama <i>et al.</i> , 2015

Conclusion

The molecules from nine medicinal plants have been reviewed. Structurally distinctive different compounds were obtained from these plants. Our study showed that medicinal plants can be a principle source of phytoconstituents as well as medicinal moieties.

References

- Hussain, M.M., Mughal, M.M.R., Alam, M.M., Dastagir, M.G., Billah, A.H.M.M. and Ismail, M. 2010. Antimicrobial activity of n-hexane and ethyl acetate extracts of *Erythrina stricta* Roxb. *Bangladesh J. Microbiol.* **27**, 65-66.
- Hussain, M.M., Tuhin, M.T.H., Akter, F. and Rashid, M.A. 2016a. Constituents of *Erythrina*- a potential source of secondary metabolites: A review. *Bangladesh Pharm. J.* **19**, 237-253.
- Hussain, M.M., Dastagir, M.G., Billah, A.H.M M. And Ismail, M. 2011. Alpinum isoflavone from *Erythrina stricta* Roxb. *Bol. Latinoam. Caribe Plant. Med. Aromat* **10**, 88-90.
- Hussain, M.M., Tahia, F. and Rashid, M.A. 2016b. Secondary metabolites from some species of *Albizia*: A review. *Bangladesh Pharm. J.* **19**, 1-8.
- Hussain, M.M., Rahman, M.S., Jabbar, A. and Rashid, M.A. 2008. Phytochemical and biological investigations of *Albizia lebbeck* Benth. *Bol. Latinoam. Caribe Plant. Med. Aromat.* **7**, 273-278.
- Yenesew, A., Akala, H.M., Twinomuhwezi, H., Chepkirui, C., Irungu, B.N., Eyase, F.L., Mugisha, M.K., Kiremire, B.T., Johnson, J.D. and Waters, N.C. 2012. The antiplasmodial and radical scavenging activities of flavonoids of *Erythrina burttii*. *Acta Tropica* **123**, 123-127.
- Yenesew, A., Midiwo, J.O., Miessner, M., Heydenreich, M. and Peter, M.G. 1998. Two prenylated flavanones from stem bark of *Erythrina burttii*. *Phytochemistry* **48**, 1439-1443.
- Yenesew, A., Irungu, B., Derese, S., Midiwo, J.O., Heydenreich, M. and Peter, M.G. 2003. Two prenylated flavonoids from the stem bark of *Erythrina burttii*. *Phytochemistry* **63**, 445-448.
- Bedane, K.G., Kusari, S., Bullach, A., Masesane, I.B., Mihigo, S.O., Spitteller, M. and Majida, R.R.T. 2017. Chemical constituents of the root bark of *Erythrina droogmansiana*. *Phytochem. Lett.* **20**, 84-88.
- Kader, M.A., Hoch, J., Berger, J.M., Evans, R., Miller, J.S., Wisse, J.H., Mamber, S.W., Dalton, J.M. and Kingston, D.G.I. 2001. *J. Nat. Prod.* **64**, 536-539.
- Runyoro, D.K.B., Joseph, C.C., Ngassapa, O.D., Darokar, M.P., Srivastava, S.K., Matee, M.I.N. and Wright, C.W. 2015. Anticandida agents from a Tanzanian plant *Albizia anthelmintica*. *J. Chin. Chem. Soc.* **62**, 1-6.
- Zhang, H., Samadi, A.K., Rao, K.V., Cohen, M.S. and Timmermann, B.N. 2011. Cytotoxic oleanane type saponins from *Albizia inundata*. *J. Nat. Prod.* **74**, 477-482.
- Carpani, G., Orsina, F., Sisti, M. and Verotta, L. 1989. Saponins from *Albizia anthelmintica*. *Phytochemistry* **28**, 863-866.
- Pertuit, D., Larshini, M., Brahim, M.A., Markouk, M., M-Offer, A-C., Paululat, T., Delemasure, S., Dutartre, P. and L-Dubois, M.A. 2017. Triterpenoid saponins from the root of *Spergularia marginata*. *Phytochemistry* **139**, 81-87.
- Vieira, P.D.B., Silva, N.L.F., Menezes, C.B., Silva, M.V.D., Silva, D.B., Lopes, N.P., Macedo, A.J., Bastida, J. andTasca, T. 2017. Trichomonocidal and parasite membrane damaging activity of bidesmosic saponins from *Manilkara rufula*. *PLOS ONE* **12**, e0188531.
- Massarani, S.M.A., Gamal, A.A.E., Halim, M.F.A.E., Said, M.S.A., Kader, M.S.A., Basudan, O.A. and Alqasoumi, S.I. 2017. New acyclic secondary metabolites from the biologically active fraction of *Albizia lebbeck*. *Saudi Pharm. J.* **25**, 110-121.
- Fang, X., Xu, X-K., Wang, G-W., Zeng, R-T., Tian, X-H., Shi, Z-R., Zhou, Z-G., Shen, Y.H. and Zhang, W-D. 2017. Guaianolide sesquiterpenoids from *Ainsliaea yunnanesis*. *Phytochemistry* **139**, 47-55
- Fotso, G.W., Kamga, J., Ngameni, B., Uesugi, S., Ohno, M., Kimura, K-I., Momma, H., Kwon, E., Furuno, H., Shiono, Y., Ingrid, S.K., Yeboah, S.O. and Ngadjui, B.T. 2017. Secondary metabolites with anti-proliferative effects from *Albizia glaberrima* var *glabrescens* Oliv. (Mimosoideae). *Nat. Product Res.* DOI: <http://dx.doi.org/10.1080/14786419.2016.1269097>.
- Runyoro, D.K.B., Srivastava, S.K., Darokar, M.P., Olipa, N.D., Joseph, C.C. and Matee, M.I.N. 2013. Anticandidiasis agents from a Tanzanian plant, *Combretum zeyheri*. *Med. Chem. Res.* **22**, 1258-1262.

- Note, O.P., Azouaou, S.A., Simo, L., Antheaume, C., Guillaume, D., Pegnyemb, D.E., Muller, C.D. and Lobstein, A. 2016. Phenotype-specific apoptosis induced by three triterpenoid saponins from *Albizzia glaberrima* (Schumach & Thonn.) Benth. *Fitoterapia* **109**, 80-86.
- Note, O.V., Jihu, D., Antheaume, C., Guillaume, D., Pegnyemb, D. E., Kilhoffer, M. C. and Lobstein, A. 2015. Triterpenoid saponins from *Albizzia boromoensis* Aubrev. & Pellegr. *Phytochemistry Lett.* **11**, 37-42.
- Simo, L.M., Note, O.P., Mbing, J.N., Aouazou, S.A., Guillaume, D., Muller, C.D., Pegnyemb, D.E. and A. Lobstein. 2017. New cytotoxic triterpenoid saponins from the roots of *Albizzia gummifera*. *Chem. Biodiversity* **14**, e1700260.
- Krief, S., Thoison, O., Sevenet, T., Wrangham, R.W. and C. Lavaud. 2005. Triterpenoid saponins anthranilates from *Albizzia grandibracteata* leaves ingested by promates in Uganda. *J. Nat. Prod.* **68**, 897-903
- Byamukama, R., Barbara, G., Namukobe, J., Heydenreich, M. and B. Kiremire. 2015. Bioactive compounds in the stem bark of *Albizzia coriaria*. *Int. J. Bio. Chem. Sci.* **9**, 1013-1024.